

RJC 2_Kinetics of Chlorella Sp Growth Models in Reducing CO2 Emissions

by Euis Nurul Hidayah

Submission date: 23-Nov-2020 12:10PM (UTC+0700)

Submission ID: 1454711439

File name: Kinerja_RJC_12_4_2019_penulis_ke-3.pdf (169.15K)

Word count: 2491

Character count: 12155

KINETICS OF *Chlorella sp* GROWTH MODELS IN REDUCING CO₂ EMISSIONS

Okik Hendriyanto Cahyonugroho^{1,*}, Dwi Dipta Yuniawati¹
and Euis Nurul Hidayah¹

¹Department of Environmental Engineering, University of Pembangunan Nasional "Veteran"
Jawa Timur, 60294, Indonesia

*E-mail: okikhc@upnjatim.ac.id

ABSTRACT

CO₂ emissions resulting from human activities are relatively higher concentrations that disrupt the equilibrium system in the air and ultimately damage the environment and human well-being. One environmentally friendly solution for CO₂ gas removal is to use microalgae (*Chlorella sp.*). *Chlorella sp.* in photobioreactors has the ability to biofixate CO₂ gas. *Chlorella sp.* was chosen because of the most numerous in freshwater and seawater. The purpose of this study was to determine the kinetic model of the effect of adding substrate concentrations and flow rate variations on the growth and development rate of *Chlorella sp.* due to exposure to pure CO₂ gas emissions. This study uses the CO₂ gas with a flow rate of 0.02 L/min, 0.04 L/min, 0.06 L/min, 0.08 L/min, 0.1 L/min and a substrate of 350 mg/L, 500 mg/L, 650 mg/L, 800 mg/L, 950 mg/L on a laboratory scale. The results showed that the most optimum growth rate occurred at the addition of substrate concentrations of 800 mg/L with a flow rate of 0.08 L/min.

Keywords: CO₂ Emission, *Chlorella sp*, Photobioreactors, Kinetic Models

© RASAYAN. All rights reserved

INTRODUCTION

The phenomenon of global warming is characterized by a gradual rise in the average temperature of the earth. Temperature changes occur in the long run slowly, but over time will be felt and have a big effect. Increased global warming due to air pollution by high CO₂. Where carbon dioxide is a colorless gas found in the atmosphere with amounts reaching 330 ppm and most comes from burning fossil fuels.¹

Research began in the 90s in anticipation of increasing flue gas globally directed to find solutions, one of which was carried out using cyanobacteria and microalgae biological activity because of its reliability in fixing CO₂ through photosynthesis reactions with the help of light energy.² Microalgae-based CO₂ fixation technology that has been widely developed is photobioreactor technology.³ Photosynthesis is generally defined as the process of forming new chemical compounds using light energy consisting of light and dark reactions. During the light reaction, the light energy is captured, whereas in the dark reaction that is the energy used to capture CO₂ from the air.

Microalgae is the most efficient plant in capturing, utilizing solar energy, and CO₂ for photosynthesis purposes.⁴ Microalgae are single-celled microscopic plants that can absorb CO₂ in an effort to reduce concentrations of CO₂ in the air.⁵ Carbon dioxide through the process of photosynthesis with the help of sunlight is converted into oxygen, so that the amount of carbon dioxide that enters the system will be reduced when it exits the microalgae system. Green cells such as leaves that contain lots of chlorophyll which are used by algae to capture light to synthesize carbon sourced from carbon dioxide (CO₂). Microalgae can fix CO₂ (10 - 50) times more efficient compared to plants.⁶ Microalgae breeding to obtain high cell density is carried out in microalgae biotechnology with optimal reactor designs and processes.⁷ *Chlorella sp.* including one group of green algae which have a structure similar to plants, one of which is a cell wall composed of cellulose. In this study, *Chlorella sp.* used because it is able to fix CO₂, rapid growth rate, as well as the production of biomass produced, is very beneficial for life.⁸

Rasayan J. Chem., 12(4), 2306-2310(2019)

<http://dx.doi.org/10.31788/RJC.2019.1245439>



Microbial growth is characterized by an increase in microbial cells due to the availability of nutrients. In the previous studies², to predict the relationship between the growth of *Chlorella Vulgaris* Buitenzorg with the substrate in the media is using a general equation model, one of which is the Monod equation model. Microbial growth can be measured by looking at the increase in biomass or the number of cells usually following a certain growth pattern in the form of a sigmoid growth curve (monod model). In order to investigate the mechanism of sorption and potential controlling steps such as mass transport, several kinetic models were tested including the pseudo-first-order kinetic model, the Elovich model and the pseudo-second-order kinetic model for a batch contact time process⁹, but the monod equation is simpler and is commonly used with the relationship of growth rate and substrate concentration.¹⁰ Kinetic studies are needed to describe the efficacy of biosorption. It is necessary to recognize the mechanism of biosorption.¹¹ Therefore this study was conducted to determine the removal of CO₂ gas by microalgae (*Chlorella sp.*) Using photobioreactor technology with the Monod equation model to determine the optimal CO₂ absorption in microalgae (*Chlorella sp.*).

EXPERIMENTAL

This research was to determine the kinetic model of the effect of adding substrate concentrations and flow rate variations on the growth and development rates of *Chlorella sp.* due to exposure to pure CO₂ gas emissions. The reactor used is a closed container with a diameter of 12 cm and a height of 18 cm with a volume of 2 liters, then the reactor is placed on a stacked 5 rack. The reactor that contains microalgae is supplied with CO₂ gas with a predetermined CO₂ flow rate for approximately 8 hours.

The running process is carried out when the reactor is ready and running based on the algal life cycle. At the time of running, algae are expected to be in a homogeneous condition. Sampling is conducted every hour during the CO₂ exposure process. After that, chlorophyll-a testing was done using a spectrophotometer to determine the biomass of microalgae in the reactor. During the running process, the reactor must be closed so that the reactor is not contaminated by external light or other foreign matter.

RESULTS AND DISCUSSION

Growth Rate (μ)

The specific growth rate values (μ) (day⁻¹) is obtained in Table-1.

Table-1: Specific Growth Rate Values (day⁻¹) on Variations in Substrate Concentration and Flow Rate

Flow Rate (L/min)	μ (day ⁻¹)				
	Substrate Concentration (mg/L)				
	350	500	650	800	950
0.02	0.8231	0.5624	0.2984	0.8031	0.3682
0.04	0.8231	0.2181	0.4004	0.379	0.6918
0.06	0.0854	0.2181	0.4004	0.379	0.6918
0.08	0.1503	0.1919	0.711	0.8375	0.6041
0.1	0.7497	0.0598	0.5207	0.302	0.3896

The table above shows that the highest coefficient of the linear regression is at the variation of flow rate 0.08 L/min for substrate concentration 800 mg/L, while the lowest coefficient value is at the variation of flow rate 0.06 L/min for substrate concentration 350 mg/L. Low μ values indicate slow microalgae growth, but low coefficient values can be influenced by cell density and differences in observation time.¹² Fluctuations in the curve can also be caused by the sampling time of each reactor so that the growth phases are not well observed. At the addition of a large substrate concentration of 950 mg/L can not produce a high growth rate, this can be due to the substrate concentration can be an inhibitor for microorganisms.

Growth Yield / Y Growth

Yield coefficient (Y) is the ratio of the amount of biomass production and the amount of substrate concentration used.¹³ By plotting the value of X_m (maximum biomass) with substrate concentration (S₀),

the results of Growth Yield (Y) can be obtained (Fig.-1) by knowing the straight-line equation with the formed slope line is the Y value as shown in Table-2.

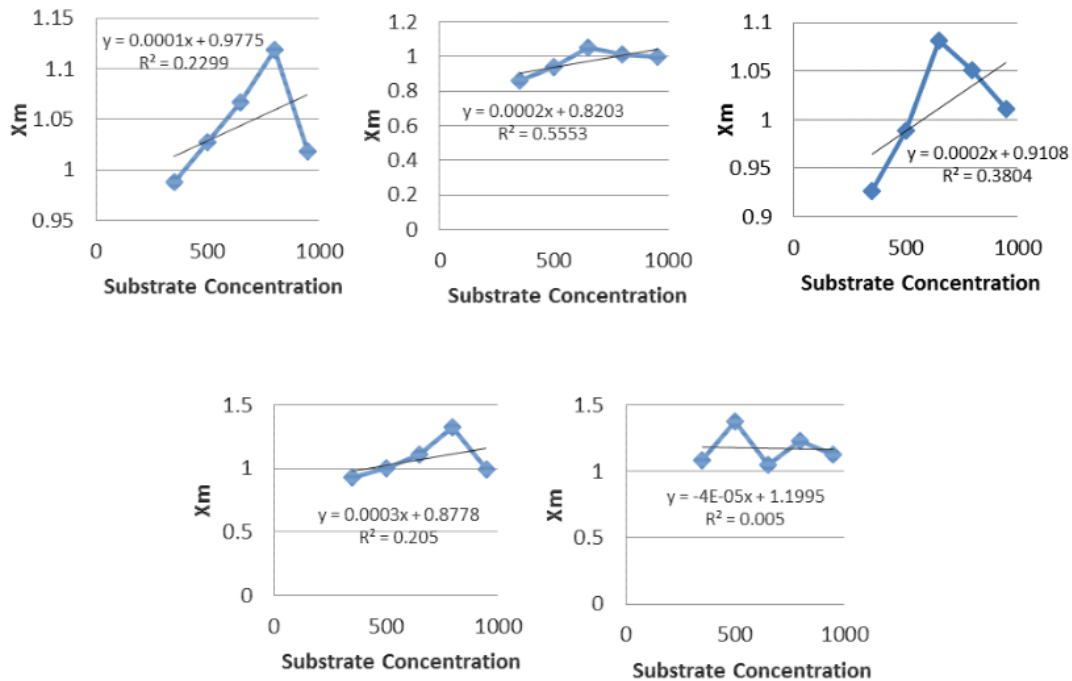


Fig.-1: The plot of Maximum Biomass (Xm) and Substrate Concentration on Flow Rate Variation.

Table-2: Growth Yield Value (Y) on Each Flow Rate Variation

Flow Rate (L/min)	Equation	Growth Yield (Y)
0.02	$X_m = 0.0001S_o + 0.9775$	0.0001
0.04	$X_m = 0.0002S_o + 0.8203$	0.0002
0.06	$X_m = 0.0002S_o + 0.9108$	0.0002
0.08	$X_m = 0.0003S_o + 0.8778$	0.0003
0.1	$X_m = 0.00004S_o + 1.1995$	0.00004

9

The results showed that the greater the CO₂ flow rates (from 0.02 L/min to 0.08 L/min), there was an increase in the value of growth yield (Y) which reached 0,0001 to 0,0003. This indicates that the time of CO₂ exposure of 0.08 L/min, there was a large conversion of new cells with an increase in chlorophyll-a concentration in algae. However, at the highest flow rate of 0.1 L/min has a small Y coefficient value which is 0.00004. This is due to the excessive concentration of gas which can be a barrier for algal growth.

Yield values indicate the amount of organic material that is converted into new cells.¹⁴ The greater the flow rate used, the smaller the coefficient of Y. A high Y value does not always indicate a better concentration of degradability because there are other factors that control the kinetics of biodegradation, including enzyme activity and initial substrate concentration¹¹.

Saturation Constants (Ks) and Maximum Growth Rate (μmax)

Ks value indicates the substrate concentration which has saturated with the growth of microorganisms. The value of Ks depends on the type of microorganisms and the type of substrate used. The maximum growth rate (μmax) is the maximum value of the growth rate at the peak point in the exponential phase

before entering the stationary phase. The values of μ_{\max} and K_s in each flow rate variation can be shown in Table-3.

Table-3: The Values of μ_{\max} and K_s in Each Flow Rate Variation

Flow rate (mg/L)	Equation	μ_{\max}	K_s
0.02	$1/\mu = -642.49 \text{ 1/S} + 3.1788$	0.315	202.117
0.04	$1/\mu = -114.12 \text{ 1/S} + 2.6748$	0.374	42.665
0.06	$1/\mu = 5554.7 \text{ 1/S} + 5.0881$	0.197	1091.704
0.08	$1/\mu = 3280.6 \text{ 1/S} + 2.4831$	0.403	1321.171
0.1	$1/\mu = 1604.1 \text{ 1/S} + 2.8718$	0.348	856.983

Table 3 shows that the optimum of μ_{\max} coefficients and K_s coefficients found in the CO_2 flow rate of 0.08 L/min, which are 0.403 and 1321.171. That is because the substrate is in maximum concentration and the growth of microalgae will experience saturation. At a flow rate of 0.1 L/min, it can be seen that the coefficient μ_{\max} and the K_s coefficient have decreased by 0.348 and 856.983. This shows that microalgae are experiencing burnout and are in a phase of decay. The greater the coefficient values of μ_{\max} and K_s indicate the concentration of the substrate will experience saturation of the growth of microalgae. K_s shows the saturation of substrate concentration on biomass growth.¹⁵ However (k_s) was unaffected by ionic strength since the reaction is taking place between charged and uncharged species.¹⁶ Based on the specific growth kinetics coefficient (μ), Growth Yield (Y), saturation constant (K_s) and maximum growth rate (μ_{\max}) show the consistency that the CO_2 flow rate of 0.08 mg/L shows the optimal coefficient value in the growth of microalgae *Chlorella sp* the CO_2 absorption process.

CONCLUSION

From the results of the research and discussion that has been described, it can be concluded that the time of exposure to CO_2 gas with each variation of CO_2 flow rate and the addition of substrate concentrations affect the growth of *Chlorella sp.* with different growth phases, but the provision of high concentrations of CO_2 exposure and high substrate concentrations can be a barrier to the growth of the algae. The most optimum value of chlorophyll-a concentration occurs at the addition of a substrate concentration of 800 mg/L with a flow rate of 0.08 L/min. Basically, the kinetic model parameters show the increasing growth rate of *Chlorella sp.* in removing CO_2 gas. Optimum CO_2 removal conditions are found at a flow rate of 0.08 L/min and substrate concentrations of 800 mg/L which have optimal coefficients of μ , Y, K_s , and μ_{\max} as well.

ACKNOWLEDGMENT

The financial support provided to this research by Research and Community Service Centre of University Pembangunan Nasional Veteran Jawa Timur (RCSC-UPN Veteran Jawa Timur), Indonesia (Contract No. SPP/95/UN.63.8/LT/V/2019) is greatly appreciated.

REFERENCES

1. A. Wijanarko, H. Hermansyah, M. Gozan, and B. A. Witarto, *Biotechnology and Bioprocess Engineering Journal*, **11(6)**, 484(2006), DOI: 10.1007/BF02932071
2. A. Wijanarko, Dianursanti, B. A. Witarto, and W. R. Soemantojo, *Makara Journal of Technology*, **8(2)**, 35(2004), DOI:10.7454/mst.v8i2.254
3. O. H. Cahyonugroho, S. N. Nindhita, *International Journal of ChemTech Research*, **11(10)**, 08(2018), DOI: 10.20902/IJCTR.2018.111002.
4. Kasrina, S. Irawati, and Wahyu, E., *Exacta Journal*, **10(1)**, 36(2012)
5. A. Mulyanto, *Indonesian Hydrosfir Journal*. **5(2)**, 13(2010)
6. B. Wang, C. Q. Lan, M. Horsman, *Biotechnol. Adv.*, **30(4)**, 904(2012) DOI:10.1016/j.biotechadv.2012.01.019
7. M. J. G. V. Barbosa, Ph.D Thesis, Wageningen University, Wageningen, The Netherland-Withsummary in Dutch, (2003).

8. N. Sopiāh, M. Adī, and S. Sīndī, *Journal of Environmental Technology*, **14(1)**, 01(2013). DOI:10.29122/jtl.v14i1.1431.
9. C. Sumithra and S. Karthikeyan, *Rasayan Journal of Chemistry*, **7(2)**, 149(2014).
10. Margono, H. Sulistyō, and Sumardi, *Technical Forum*, pp. 65-71,(2001).
11. P. P. Kumar, Y. P. Kumar, and B. V. Rao, *Rasayan Journal of Chemistry*, **10(3)**, 1025(2017), DOI: 10.7324/RJC.2017.1031683
12. Fahria, Thesis, Department of Environmental Engineering, Faculty of Civil Engineering and Planning, National Development University, Surabaya, (2012),
13. Metcalf and Eddy, Inc., (1979), *Wastewater Engineering Treatment Disposal Reuse*, Mc Graw Hill, Inc., New York. pp. 234-246.
14. Wahyu, S. Istiyantyo, C. K. Andri and Silviana, In *Proceedings of the National Seminar on Chemical Engineering*. Bandung, Indonesia, pp. 143-150 (2014).
15. D. W. Septiani, A. Slamet, and J. Hermana, *Pomits Engineering Journal*, **15(3)**, 20(2014)
16. Salem Et. Ashoor, Fatma M. Ashoor and Noori Y. Salman, *Rasayan Journal of Chemistry*, **6(1)**, 1 (2013).

[RJC-5439/2019]

RJC 2_Kinetics of Chlorella Sp Growth Models in Reducing CO2 Emissions

ORIGINALITY REPORT

9%

SIMILARITY INDEX

4%

INTERNET SOURCES

7%

PUBLICATIONS

3%

STUDENT PAPERS

PRIMARY SOURCES

1	downloads.hindawi.com Internet Source	2%
2	Submitted to Ocean County College Student Paper	1%
3	Submitted to Bridgepoint Education Student Paper	1%
4	etd.auburn.edu Internet Source	1%
5	EMAN S. H. KHALED. "Kinetics and Mechanism of Oxidation of a Ternary Complex Involving Dipicolinatochromium(III) and Proline by N-bromsuccinimide", <i>BioInorganic Reaction Mechanisms</i> , 2006 Publication	1%
6	Ragil Adi Prasetyo, Ambar Pertiwiningrum, Yuny Erwanto, Lies Mira Yusiati, Nanung Agus Fitriyanto. "Chapter 19 Characterization of Pseudomonas sp. LS3K as Nitrate Removal	1%

Agent at Different C/N Ratios Under Aerobic Condition", Springer Science and Business Media LLC, 2018

Publication

7

G. Yue, A. Fadl, T. Barek, Z. Zhang, J. Major. "Experimental Study of Aerosol Deposition in Pulsating Balloon Structures", Inhalation Toxicology, 2009

Publication

1%

8

czkj.ci.cqvip.com

Internet Source

1%

9

www.freepatentsonline.com

Internet Source

<1%

10

"Proceeding of the 2nd International Conference on Tropical Agriculture", Springer Science and Business Media LLC, 2018

Publication

<1%

11

pdfs.semanticscholar.org

Internet Source

<1%

12

Stanley I. Pinel, Hans R. Signer, Erwin V. Zaretsky. "Comparison Between Oil-Mist and Oil-Jet Lubrication of High-Speed, Small-Bore, Angular-Contact Ball Bearings", Tribology Transactions, 2001

Publication

<1%

Exclude quotes Off

Exclude matches Off

Exclude bibliography On